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Localization of Transgenes for Drosophila Models of Myotonic Dystrophy Type 1

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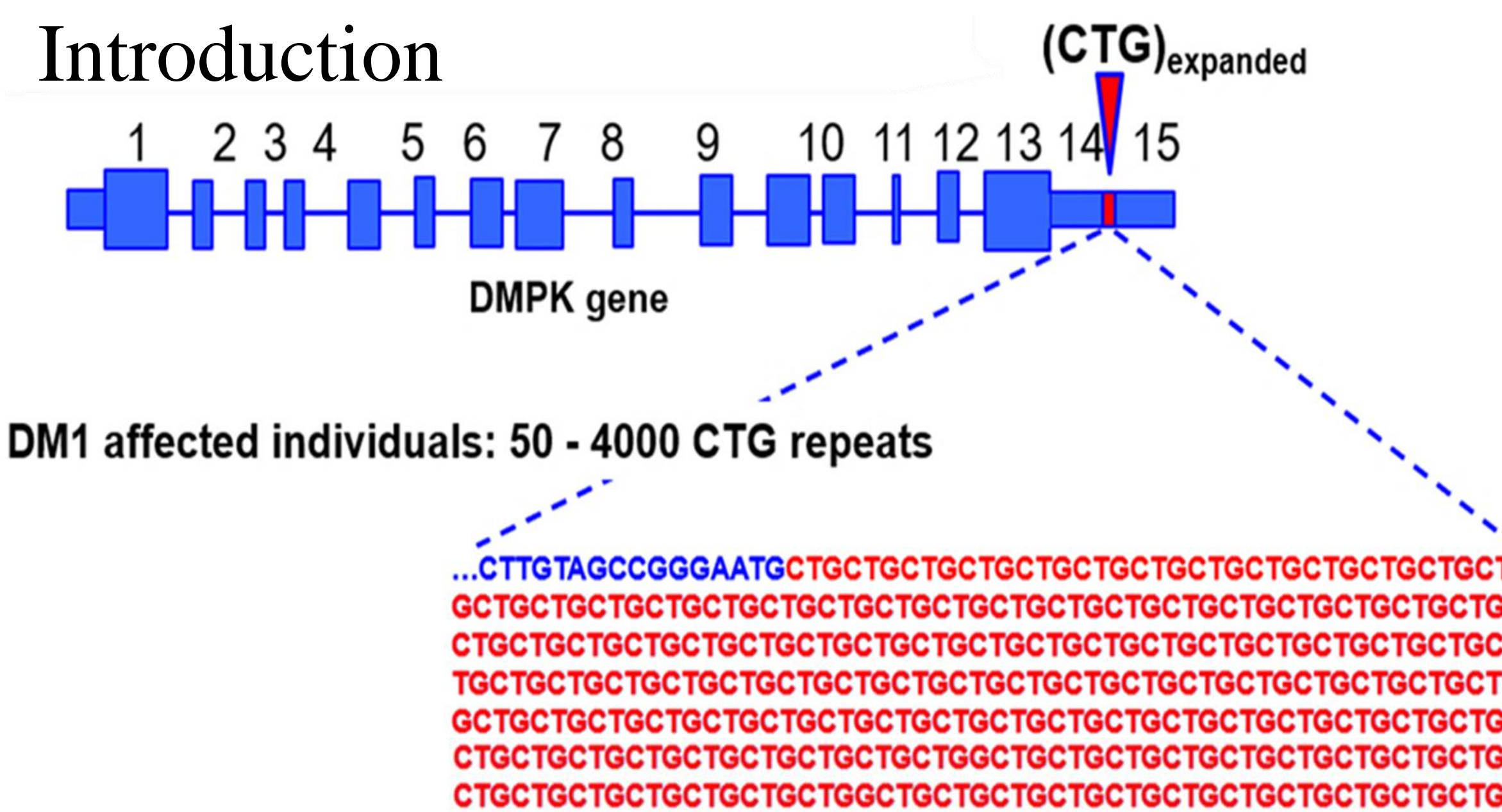
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Introduction

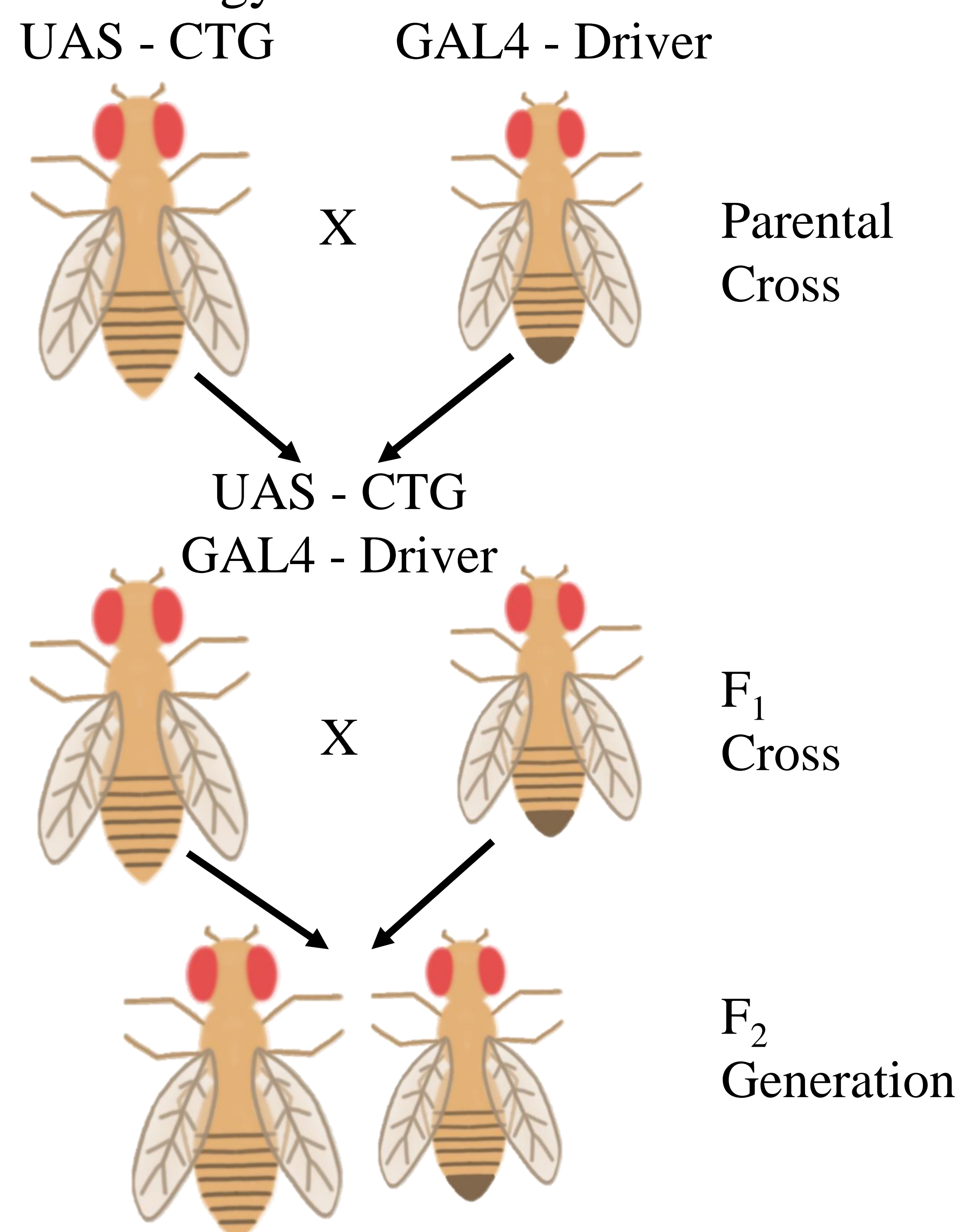


- Increased number of repeats has increased severity.

Drosophila in DM1 Research

- Transgenic DM1 *Drosophila* lines contain either 60, 250, or 480 CTG repeats.^{1,2}
- Many of the driver *Drosophila* lines have not been used to express DM1 transgenes previously.

Methodology



Phenotypes that show the ratios in F₂ Generation:

- Eye color
- Climbing capabilities³ and flight capabilities⁴
 - Shown to be deficient in DM1 expressing flies
- Skeletal muscle density

Genetic Crosses

Image (right) of muscle sections comparing muscle loss of DM1 and severe DM1 to control (MHC-GAL4). Image from Garcia-Lopez *et al. PLoS One*. 2008.

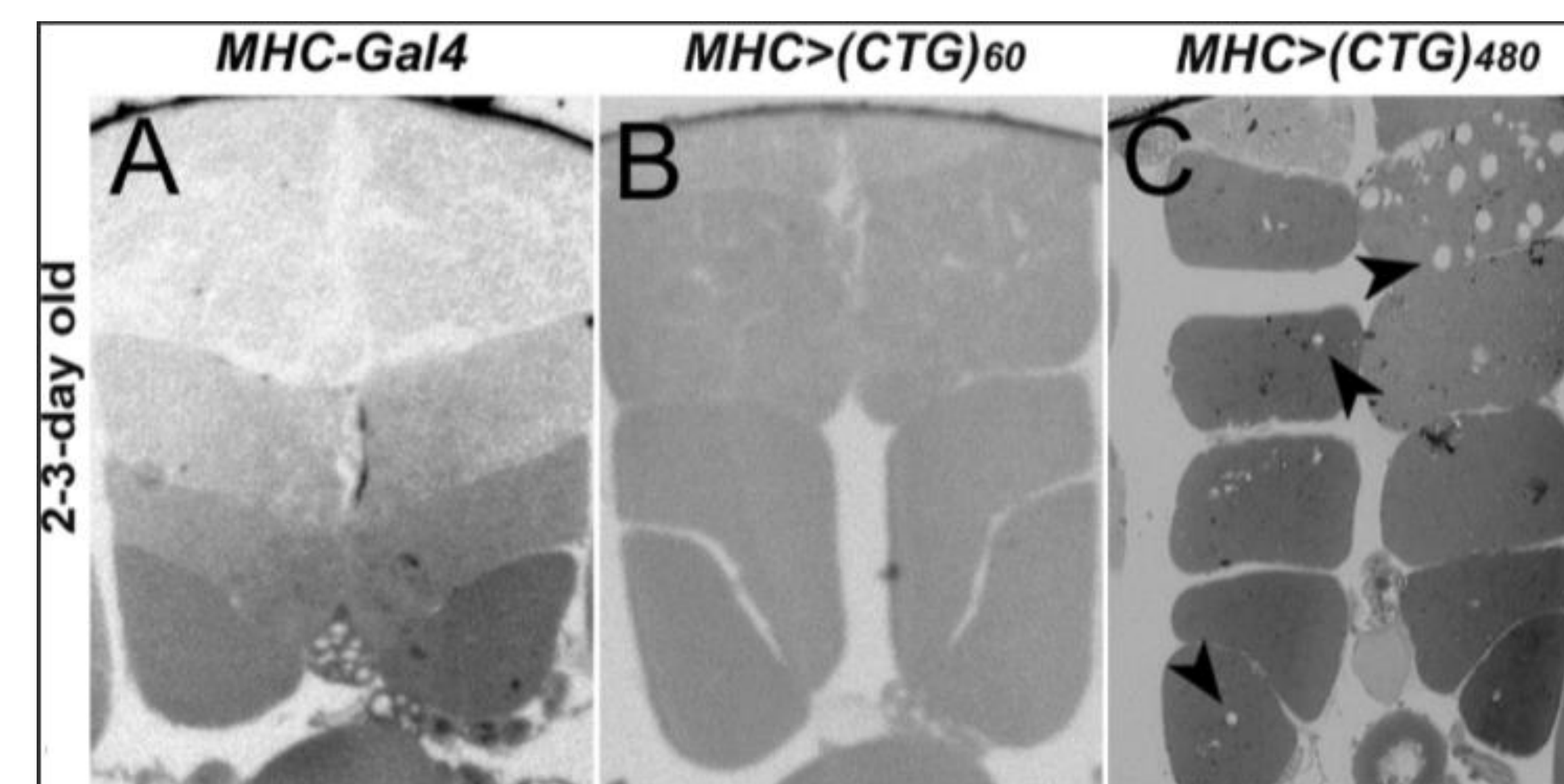


Image (left) of salivary gland dissection from 3rd instar larvae.

Climbing Assay

Flight Assay

Muscle Histology



Isolate and purify DNA from 3rd instar larvae from each line

Design primers for long range PCR and create probe for transgenes and chromosome 1, 2, and 3 via nick translation

Dissection of 3rd instar larvae

Fluorescent *in situ* hybridization of
transgene and chromosome probes
to isolated chromosomes and
counterstaining with Hoechst stain

Observe fluorescence of probes
under microscope

Chromosome 2

Chromosome 3

Example (above) of what could be seen after FISH is conducted with CTG₂₅₀ labeling, Chromosome 2 (left) possess two fluorescent bands indicating presence of transgene (pink) and the chromosome specific band (orange), as opposed to chromosome 3 (right) that shows only the chromosome-specific fluorescent band (blue).

Results

The presence of white eyed flies in the F₂ progeny indicate chromosomal location.

Cross: UAS – CTG x Hsp70-Gal4 (on the 2nd chromosome)

- Present
 - (CTG)₆₀
 - i(CTG)₄₈₀
- Absence
 - (CTG)₂₅₀

Conclusions

The preliminary results suggest:

- (CTG)₂₅₀ is likely localized to chromosome 2.
- i(CTG)₄₈₀ and (CTG)₆₀ are not likely localized to chromosome 2.

Future Research

This project is ongoing. After classical genetics indicate chromosomal location, fluorescent *in situ* hybridization (FISH) of polytene chromosome preparations will be used to narrow down their location on the chromosome. Then PCR and sequencing will be used to verify the location. This information will be used to assess contributions of signaling pathways in DM1.

References

1. Cerro-Herreros E, Chakraborty M, Pérez-Alonso M, Artero R, Llamusi B. Expanded CCUG repeat RNA expression in *Drosophila* heart and muscle trigger Myotonic Dystrophy type 1-like phenotypes and activate autophagocytosis genes. Scientific Reports. 2017;7(1):2843. doi:10.1038/s41598-017-02829-3
2. Garcia-Lopez A, Monferrer L, Garcia-Alcover I, Vicente-Crespo M, Alvarez-Abril MC, Artero RD. Genetic and Chemical Modifiers of a CUG Toxicity Model in *Drosophila*. PLoS ONE. 2008;3(2):e1595.
<https://www.ncbi.nlm.nih.gov/pmc/articles/PMC2220037/>. doi:10.1371/journal.pone.0001595
3. de Vries SEJ, Clandinin T. Optogenetic stimulation of escape behavior in *Drosophila melanogaster*. Journal of Visualized Experiments: JoVE. 2013;(71):50192. doi:10.3791/50192
4. Babcock, D., Gantezky, B. An Improved Method for Accurate and Rapid Measurement of Flight Performance in *Drosophila*. Journal of Visualized . 2014;84. <http://www.jove.com/video/51223>. doi:10.3791/51223.

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For Further Information

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